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- (73) Proprietor: **THE ROCKEFELLER UNIVERSITY**
1230 York Avenue
New York, NY 10021 (US)
(72) Inventor: **Cerami, Anthony**
Ram Island Drive
Shelter Island New York 11964 (US)
Inventor: **Ulrich, Peter**
500 East 63rd Street
New York New York 10021 (US)
Inventor: **Brownlee, Michael**
500 East 63rd Street
New York New York 10021 (US)

- (74) Representative: **Denmark, James et al**
Bailey, Walsh & Co.
5 York Place
Leeds LS1 2SD Yorkshire (GB)

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Description

The Applicants are co-authors of the following articles directed to the subject matter of the present invention: "COVALENT ATTACHMENT OF SOLUBLE PROTEINS BY NONENZYMATICALLY GLYCOSYLATED COLLAGEN; ROLE IN THE IN SITU FORMATION OF IMMUNE COMPLEXES, Brownlee, M., Pongor, S., Cerami, A., *J. Exp. Med.*, 158, pp. 1739-1744 (1983); and "AGING OF PROTEINS: ISOLATION AND IDENTIFICATION OF A FLUORESCENT CHROMOPHORE FROM THE REACTION OF POLYPEPTIDES WITH GLUCOSE", Pongor, S., Ulrich, P., Bencsath, A. A., and Cerami, A., *Proc. Natl. Acad. Sci. USA*, 81, pp. 2682-2688 (1984).

The present invention relates generally to the aging of proteins resulting from reaction of glucose, and particularly to the nonenzymatic glycosylation of proteins and subsequent reactions leading to advanced glycosylation end products, and to methods and agents for their inhibition.

The reaction between glucose and proteins has been known for some time. Its earliest manifestation was in the appearance of brown pigments during the cooking of food, which was identified by Maillard in 1912, who observed that glucose or other reducing sugars react with amino acids to form adducts that undergo a series of dehydrations and rearrangements to form stable brown pigments, Maillard, L.D. *C.R. Acad. Sci.*, 154, pp. 66-68 (1912).

In the years that followed the initial discovery by Maillard, food chemists studied the hypothesized reaction in detail and determined that stored and heat treated foods undergo nonenzymatic browning as a result of the reaction between glucose and the polypeptide chain, and that the proteins are resultingly crosslinked and correspondingly exhibit decreased bioavailability. At this point, it was determined that the pigments responsible for the development of the brown colour that develops as a result of protein glycosylation possessed characteristic spectra and fluorescent properties; however, the chemical structure of the pigments had not been specifically elucidated.

The reaction between reducing sugars and food proteins discussed above was found in recent years to have its parallel *in vivo*. Thus, the nonenzymatic reaction between glucose and the free amino groups on proteins to form a stable amino, 1-deoxy ketosyl adduct, known as the Amadori product, has been shown to occur with hemoglobin, wherein a rearrangement of the amino terminal of the β -chain of hemoglobin by reaction with glucose, forms the adduct known as hemoglobin A_{1c}. The reaction has also been found to occur with a variety of other body proteins, such as lens crystal-

lins, collagen and nerve proteins. See Bunn, H.F., Haney, D.N., Gabbay, K.H. and Gallop, P.H., *Biochem. Biophys. Res. Comm.*, 67, pp. 103-109 (1975); Koenig, R.J., Blobstein, S.H. and Cerami, A., *J. Biol. Chem.*, 252, pp. 2992-2997 (1975); Monnier, V.M. and Cerami, A. in *Maillard Reaction in Food and Nutrition*, ed. Waller, G.A., American Chemical Society, 215, pp. 431-448 (1983); and Monnier, V.M. and Cerami, A., *Clinics in Endocrinology and Metabolism*, 11, pp. 431-452 (1982).

Moreover, brown pigments with spectral and fluorescent properties similar to those of late-stage Maillard products have also been observed *in vivo* in association with several long-lived proteins, such as lens proteins and collagen from aged individuals. An age-related linear increase in pigment was observed in human dura collagen between the ages of 20 to 90 years. See Monnier, V.M. and Cerami, A., *Science*, 211, pp. 491-493 (1981); Monnier, V.M. and Cerami, A., *Biochem. Biophys. Acta*, 760, pp. 97-103 (1983); and Monnier, V.M., Kohn, R.R. and Cerami, A., "Accelerated Age-Related Browning of Human Collagen in Diabetes Mellitus", *Proc. Natl. Acad. Sci.*, 81, pp. 583-587 (1984). Interestingly, the aging of collagen can be mimicked *in vitro* by the crosslinking induced by glucose; and the capture of other proteins and the formation of adducts by collagen, also noted, is theorized to occur by a crosslinking reaction, and is believed to account for the observed accumulation of albumin and antibodies in kidney basement membrane. See Brownlee, M., Pongor, S. and Cerami, A., *J. Exp. Med.*, 158, pp. 1739-1744 (1983); and Kohn, R. R., Cerami, A. and Monnier, V.M., *Diabetes*, 33(1), pp. 57-59 (1984).

In parent Application Serial No. 590,820 (now U.S. Patent No. 4,665,192) and in Pongor, S.M., et al., *supra*, a fluorescent chromophore was isolated and identified which was found to be present in certain browned polypeptides such as bovine serum albumin and poly-L-lysine, and was assigned the structure 2-(2-furoyl)-4(5)-2(furanyl)-1H-imidazole. The compound was found to exist in a tautomeric state and has incorporated in its structure two peptide-derived amine nitrogens. The incorporation of these amine nitrogens and two glucose residues in the compound suggested that its peptide-bound precursor may be implicated in the *in vivo* crosslinking of proteins by glucose, which is observed in the late stage of the Maillard process. [See Chang, J.C.F., Ulrich, P.C., Bucala, R. and Cerami, A., *J. Biol. Chem.*, 260, pp. 7970-7974 (1985)]. This chromophore made possible the identification of the advanced glycosylation end products and assisted additional investigations seeking to clarify the protein aging process and to identify the specific chemistry involved in order to develop methods and agents for its inhibition. It is to this

purpose that the present Application is directed.

In accordance with the present invention, a method and associated agents are disclosed for the inhibition of protein aging. In particular, agents for inhibiting protein aging due to the formation of advanced glycosylation end products may be selected from those materials capable of reacting with the early glycosylation product from the reaction of glucose with proteins and preventing further reactions. Thus, for example, compounds or compositions having active nitrogen-containing substituents such as hydrazine groups, have been theorized to be suitable, and compounds such as aminoguanidine and α -hydrazinohistidine have been found to be suitable. These agents appear to react with the early glycosylation product at its reactive carbonyl and thereby prevent the same from later forming the advanced glycosylation end products which lead to protein crosslinks.

The present invention also relates to a method for inhibiting protein aging by contacting the initially glycosylated protein at the stage of the early glycosylation product with a quantity of one or more of the agents of the present invention. In the instance where the present method has industrial application, one or more of the agents may be applied to the proteins in question, either by introduction into a mixture of the same in the instance of a protein extract, or by application or introduction into foodstuffs containing the protein or proteins, all to prevent premature aging and spoilage of the particular foodstuffs.

In the instance where the present method has therapeutic application, the animal host intended for treatment may have administered to it a quantity of one or more of the agents, in a suitable pharmaceutical form. Administration may be accomplished by known techniques, such as oral, topical and parenteral techniques such as intradermal, subcutaneous, intravenous or intraperitoneal injection, as well as by other conventional means. Administration of the agents may take place over an extended period of time at a dosage level of, for example, up to about 25 mg/kg.

The ability to inhibit the formation of advanced glycosylation end products carries with it significant implications in all applications where protein aging is a serious detriment. Thus, in the area of food technology, the retardation of food spoilage would confer an obvious economic and social benefit by making certain foods of marginal stability less perishable and therefore more available for consumers. Spoilage would be reduced as would the expense of inspection, removal and replacement, and the extended availability of the foods could aid in stabilizing their price in the marketplace. Similarly, in other industrial applications where the perishability of proteins is a problem, the admixture of the

agents of the present invention in compositions containing such proteins would facilitate the extended useful life of the same. Presently used food preservatives and discoloration preventatives such as sulfur dioxide, known to cause toxicity including allergy and asthma in animals, might be replaced with compounds such as those described herein.

The present method has particular therapeutic application as the Maillard process acutely affects several of the significant protein masses in the body, among them collagen, elastin, lens proteins, and the kidney glomerular basement membranes. These proteins deteriorate both with age (hence the application of the term "protein aging") and as one of the sequelae of diabetes. Consequently, the ability to either retard or substantially inhibit the formation of advanced glycosylation end products carries the promise of favourably treating a significant adverse effect of diabetes and, of course, improving the quality and perhaps duration of animal life.

Accordingly, it is a principal object of the present invention to provide a method for inhibiting the extensive cross-linking of proteins that occurs as an ultimate consequence of the reaction of the proteins with glucose or other reducing sugars, by correspondingly inhibiting the formation of advanced glycosylation end products.

It is a further object of the present invention to provide a method as aforesaid which is characterised by a reaction with early glycosylation products.

It is a further object of the present invention to provide a method as aforesaid which prevents the rearrangement and cross-linking of the said early glycosylation products to form the said advanced glycosylation end products.

It is a yet further object of the present invention to provide agents capable of participating in the reaction with the said early glycosylation products in the method as aforesaid.

It is a still further object of the present invention to provide therapeutic methods of treating the adverse consequences of protein aging, manifest in the embrittlement of animal protein and the browning and spoilage of foodstuffs.

Other objects and advantages will become apparent to those skilled in the art from a consideration of the ensuing description which proceeds with reference to the following illustrative drawings.

FIGURE 1 is a graph showing the results of studies aimed at inhibiting the formation of advanced glycosylation end products in albumin which had been reacted with a quantity of glucose, on an *in vitro* basis.

FIGURE 2 is a graph showing the results of studies aimed at inhibiting protein entrapment and accumulation by glycosylated structural proteins

such as collagen.

FIGURE 3A is a graph of the degree of solubilization of collagen incubated with glucose, with and without treatment with an agent of the present invention.

FIGURE 3B is a photograph of a polyacrylamide gel showing separation of protein fragments after cyanogen bromide digestion of collagen incubated with glucose with and without an agent of the present invention.

FIGURE 4 is a graph of the results of an *in vivo* study examining the extent of protein bound to the glomerular basement membrane of diabetic rats to certain of which an agent of the present invention had been administered.

In accordance with the present invention, compositions and associated methods have been developed which inhibit the formation of advanced glycosylation end products in a number of target proteins existing in both animals and plant material. In particular, the invention relates to compositions which may contain one or more agents that are capable of inhibiting the formation of advanced glycosylation end products on such target proteins, by reacting with the carbonyl moiety of the early glycosylation product that is formed by the initial glycosylation of the protein.

It is the carbonyl group located near the junction between sugar and protein segments of the early glycosylation product that is theorized to comprise an active site that causes the further cross-linking of the protein to form the advanced glycosylation end product, and likewise contributes to the entrapment of other proteins that is evident in the development *in vivo* of conditions such as skin wrinkling, certain kidney diseases, atherosclerosis, osteoarthritis and the like. Similarly, plant and animal material that undergoes nonenzymatic browning deteriorates and, in the case of food-stuffs, become spoiled or toughened and consequently, inedible. Thus, the reaction of the compounds of this invention with this carbonyl moiety is believed to inhibit the late stage Maillard.

The rationale of the invention is to use agents which block the post-glycosylation step, i.e. the formation of fluorescent chromophores such as that identified in Pongor et al., *supra* and Farmer et al., *supra*, among others, whose presence is associated with, and leads to, the adverse sequelae of diabetes and aging. An ideal agent would prevent the formation of a chromophore and its associated cross-links of proteins to proteins and trapping of proteins on other proteins, such as that which occurs in arteries and in the kidney.

The present invention does not attempt to prevent initial protein glycosylation, as it would be nearly impossible to use agents which prevent the reaction of glucose with protein amino groups. The

agents that are capable of preventing initial glycosylation are likely to be highly toxic, and since the initial glycosylation comes to equilibrium in about three weeks, there is inadequate time available to achieve this objective. Instead, the ideal agent would prevent or inhibit the long-term, post-glycosylation steps that lead to the formation of the ultimate advanced glycosylation end products that are a direct cause of the pathology associated with aging and diabetes.

The chemical nature of the early glycosylation products with which the compounds of the present invention is believed to react are speculative. Early glycosylation products with carbonyl moieties which are involved in the formation of advanced glycosylation products, and which may be blocked by reaction with the compounds of the present invention, have been postulated. In one case, the reactive carbonyl moieties of Amadori products or their further dehydration and/or rearrangement products, may condense to form advanced glycosylation endproducts. Another proposed mechanism is the formation of reactive carbonyl compounds, containing one or more carbonyl moieties (such as glycoaldehyde or 3-deoxyglucosone) from the cleavage of Amadori or other early glycosylation products (see, for example, Gottschalk, A. (1972) in *The Glycoproteins* (Gottschalk, A., ed) Part A, pp. 141-157, Elsevier Publishing Co., New York; Reynolds, T.M. (1965) *Adv. Food Res.*, 14, pp. 167-283), and by subsequent reactions with an amine or Amadori product to form carbonyl containing advanced glycosylation products such as the alkylformylglycosylpyrroles described by Farmer et al., *supra*.

Several investigators have studied the mechanism of advanced glycosylation product formation. *In vitro* studies by Eble et al., (1983), "Nonenzymatic Glucosylation and Glucose-dependent Cross-linking of Protein", *J. BIOL. CHEM.*, 258:9406-9412, concerned the crosslinking of glycosylated protein with nonglycosylated protein in the absence of glucose. Eble et al. sought to elucidate the mechanism of the Maillard reaction and accordingly conducted controlled initial glycosylation of RNAase as a model system, which was then examined under varying conditions.

In one aspect, the glycosylated protein material was isolated and placed in a glucose-free environment and thereby observed to determine the extent of cross-linking. Eble et al. thereby observed that cross-linking continued to occur not only with the glycosylated protein but with non-glycosylated proteins as well. One of the observations noted by Eble et al. was that the reaction between glucose and the protein material appeared to occur at the location on the protein chain of the amino acid lysine. Confirmatory experimentation conducted by

Eble et al. in this connection demonstrated that free lysine would compete with the lysine on RNAase for the binding of glucose. Thus, it might be inferred from these data that lysine may serve as an inhibitor of advanced glycosylation; however, this conclusion and the underlying observations leading to it should be taken in the relatively limited context of the model system prepared and examined by Eble et al.

The experiments of Eble et al. do not suggest the reactive cleavage product mechanism or any other mechanism in the *in vivo* formation of advanced glycosylation endproducts in which glucose is always present. The use of lysine as an inhibitor in the Eble et al. model system has no bearing upon the utility of the compounds of the present invention in the inhibition of advanced glycosylated endproducts formation in the presence of glucose *in vivo* and the amelioration of complications of diabetes and aging.

Accordingly, the compositions useful in the present invention comprise or contain agents capable of reacting with the active carbonyl intermediate of the early glycosylation product. Suitable agents include compounds having an active nitrogen-containing group or substituent such as a hydrazine group. Also, the agent of compound may be at least partially derived from an amino acid, including the esters and amides thereof, as compounds having this derivation are generally biocompatible with the target proteins to be contacted. For example, the agent may comprise a compound such as aminoguanidine and α -hydrazinohistidine, or mixtures of these agents or compounds. Each of these agents or compounds possesses an active nitrogen-containing substituent that is believed to react with the carbonyl of the early glycosylation product. Consequently, reaction of the agents with the glycosyl-lysine moiety of a protein would prevent this moiety from forming crosslinks with other groups.

Hollis and Strickberger (*Diabetologia* 28:282-5 [1985]) found that *in vivo* administration of the compound α -hydrazinohistidine, a known inhibitor of the enzyme histidine decarboxylase, reduces the accumulation of albumin in the aortas of rats. The authors proposed that the drug acted to reduce production of histamine in this tissue, and that histamine is therefore the mediator of low density lipoprotein accumulation which is implicated in atherosclerotic disease. In contrast, aminoguanidine is known to increase levels of histamine (See Lindberg et al., *Acta Obst. Gynecol. Scandinav.*, 45: 131-139, (1966)) and α -hydrazinohistidine and aminoguanidine therefore have opposing effects on histamine levels. It can therefore be seen that the present findings that both α -hydrazinohistidine and aminoguanidine have efficacy *in vivo* and *in vitro* to

reduce protein crosslinking rules out from consideration and consequently distinguishes the mechanism proposed by Hollis and Strickberger as the explanation of the manner in which the compounds of the present invention might work to reduce advanced glycosylation end product formation. Moreover, the findings of Hollis and Strickberger are distinguishable from the concept and application of the present invention because the mechanism of histamine synthesis suppression by α -hydrazinohistidine suggested by the authors is functionally distinct from the underlying concept of the present invention, and it is believed, may even be placed in question by the latter.

Thus, the agents of the present invention have been identified and tested on the basis of their ability to react with the carbonyl moiety of the early glycosylation product to form a highly stable adduct, and would not have been suggested from the work of Hollis and Strickberger.

The compound aminoguanidine is known to have low toxicity in animals. According to the 1978 Registry of Toxic Effects of Chemical Substances, aminoguanidine base has a LD₅₀ when administered subcutaneously of 1258 mg/kg in rats and 963 mg/kg in mice. The hydrochloride derivative had a LD₅₀ in rats of 2984 mg/kg when given subcutaneously. The latter compound exhibits very low toxicity.

In the instance where the composition of the present invention is utilized to manufacture a pharmaceutical for *in vivo* or therapeutic purposes, it may be noted that the compounds or agents used therein are biocompatible. Pharmaceutical compositions may be prepared with a pharmaceutically effective quantity of the agents or compounds of the present invention and may include a pharmaceutically acceptable carrier, selected from known materials utilised for this purpose. Such compositions may be prepared in a variety of forms, depending on the method of administration. For example, aminoguanidine may be derivatized to the hydrochloride salt from the commercially available bicarbonate salt to improve its solubility and to make it less irritating for intraperitoneal injection. Also, a liquid form would be utilised in the instance where administration is by intravenous or intraperitoneal injection, while, if appropriate, tablets, capsules, etc., may be prepared for oral administration. For application to the skin, a lotion or ointment may be formulated with the agent in a suitable vehicle, perhaps including a carrier to aid in penetration into the skin. Other suitable forms for administration to other body tissues are also contemplated.

The present invention likewise relates to methods for inhibiting the formation of advanced glycosylation end products *in vitro*, which comprise

contacting the target proteins with the composition of the present invention. In the instance where the target proteins are contained in foodstuffs, whether plant or animal origin, these foodstuffs could have applied to them by various conventional means a composition containing the present agents. Likewise, in the instance where therapeutic applications are intended, the animals to be treated would have administered to them a regular quantity of the pharmaceutical composition of the present invention. Administration could take place, for example, daily, and an effective quantity of the agent or compound of the present invention could range up to 25 mg/kg of body weight of the animal. A topical preparation may, for example, include up to 10% of the agent or composition in an ointment or lotion for application to the skin. Naturally, some variation in these amounts is possible, and the suggested amounts are provided in fulfillment of applicants' duty to disclose the best mode for the practice of the present invention.

As is apparent from a discussion of the environment of the present invention, the present methods and compositions hold the promise for arresting the aging of key proteins both in animals and plants, and concomitantly, conferring both economic and medical benefits as a result thereof. In the instance of foodstuffs, the administration of the present composition holds the promise for retarding food spoilage thereby making foodstuffs of increased shelf life and greater availability to consumers. Replacement of currently-used preservatives, such as sulfur dioxide known to cause allergies and asthma in humans, with non-toxic, biocompatible compounds is a further advantage of the present invention.

The therapeutic implications of the present invention relate to the arrest of the aging process which has, as indicated earlier, been identified in the aging of key proteins by advanced glycosylation and crosslinking. Thus, body proteins, and particularly structural body proteins such as collagen, elastin, lens proteins, nerve proteins and kidney glomerular basement membranes would all benefit in their longevity and operation from the practice of the present invention. The present invention thus reduces the incidence of pathologies involving the entrapment of proteins by crosslinked target proteins, such as retinopathy, cataracts, diabetic kidney disease, glomerulosclerosis, peripheral vascular disease, arteriosclerosis obliterans, peripheral neuropathy, stroke, hypertension, atherosclerosis, osteoarthritis, periarticular rigidity, loss of elasticity and wrinkling of skin, stiffening of joints, glomerulonephritis, etc. Likewise, all of these conditions are in evidence in patients afflicted with diabetes mellitus. Thus, the present therapeutic method is relevant to treatment of the noted con-

ditions in patients either of advanced age or those suffering from one of the mentioned pathologies.

Protein crosslinking through advanced glycosylation product formation can decrease solubility of structural proteins such as collagen in vessel walls [see Brownlee et al., *Science*, 232, pp. 1629-1632, (1986)], as well as trap serum proteins, such as lipoproteins, to the collagen. Also, this may result in covalent trapping of extravasated plasma proteins in subendothelial matrix, and reduction in susceptibility of both plasma and matrix proteins to physiologic degradation by enzymes. (See Brownlee et al., *Diabetes*, 35, Suppl. 1, p. 42A (1986).) For these reasons, the progressive occlusion of diabetic vessels induced by chronic hyperglycemia has been hypothesized to result from excessive formation of glucose-derived crosslinks. Such diabetic macrovascular changes and microvascular occlusion can be effectively prevented by chemical inhibition of advanced glycosylation product formation utilizing a composition and the methods of the present invention.

Diabetes-induced changes in the deformability of red blood cells, leading to more rigid cell membranes, is another manifestation of cross-linking and aminoguanidine has been shown to prevent it *in vivo*.

A further consequence of diabetes is the effect of hyperglycemia on bone matrix differentiation resulting in decreased bone formation usually associated with chronic diabetes. In animal models, diabetes reduces matrix-induced bone differentiation by 70% and *in vitro* models mimic this effect. Aminoguanidine prevents the decreased bone formation in this model.

The present invention will be better understood from a consideration of the following illustrative examples, reviewing the selection and testing of certain of the agents of the present invention on both an *in vitro* and *in vivo* basis.

EXAMPLE 1

To measure the ability of certain of the agents of the present invention to inhibit the production of advanced glycosylation end products *in vitro*, albumin and glucose were incubated together for two weeks in the presence of several test agents. Samples were taken at regular intervals for analysis. Advanced glycosylation end products were measured as appearance of fluorescent compounds, and early glycosylation products were measured by incorporation of radiolabeled glucose into albumin. Reaction conditions were as follows. Each mixture contained 6 mg/mL bovine serum albumin, 200 mM glucose, 200 mM test agent (either aminoguanidine hydrochloride, α -hydrazinohistidine, or lysine), and approximately 9.5×10^6

counts per minute of ^{14}C -glucose in 0.5M phosphate buffer, pH 7.6. The radiolabeled glucose was prepurified before use to eliminate breakdown products which might react with the albumin and lead to an erroneous indication of the degree of early glycosylation product formation. Reaction mixtures were incubated at 37°C , and samples were taken after 0.5, 1.0, 1.5, and 2 weeks. Control mixtures lacked glucose or agent.

After the incubation periods, samples were treated as follows. After dialysis to remove all unbound glucose, the amount of protein present was measured with a standard dye-binding assay. The amount of glucose which became bound to the albumin, a measure of early glycosylation products, was determined by precipitating albumin with trichloroacetic acid and measuring the radioactivity of the bound glucose using scintillation counting. Advanced glycosylation end products were quantitated by determining the fluorescence of the albumin described in parent EP Patent No. 0175764 and as described by Pongor et al., *supra*. Spectral measurements on excitation and emission maxima were made on all samples to ensure that these values had not been shifted as a result of adduct formation with inhibitors.

The result of this experiment are expressed graphically in Figure 1. For each sample, incorporation of radiolabeled glucose is indicated by the solid portion of the bar, and fluorescence is indicated in the open portion of the bar.

All values are expressed as per milligram of albumin. In all further discussions, aminoguanidine refers to the hydrochloride derivative.

The results show that the glucose and albumin react to form a large amount of fluorescent advanced glycosylation end products after 0.5, 1, 1.5 and 2 weeks of incubation ("GLUCOSE + BSA"). Inclusion of 200 mM aminoguanidine dramatically reduced by as much as eightfold with formation of fluorescent compounds, by comparison with the control samples after a two-week incubation ("BSA + GLUCOSE + I#2). Inclusion of 200 mM α -hydrazinohistidine also reduced formation of advanced glycosylation end products as measured by fluorescence ("BSA + GLUCOSE + I#1). Lysine appeared to cause an increase in fluorescent compound formation ("BSA + GLUCOSE + LYS"), but as will be seen in the next experiment, it had the ability to reduce protein crosslinking. The amount of early glycosylation end products, as measured by glucose incorporation, was nearly unchanged in all reactions. The control incubation without glucose showed little development of fluorescent products (A).

These results show that aminoguanidine, and to a lesser extent, α -hydrazinohistidine, reduce the formation of fluorescent compounds when glucose

and albumin react over time, and indicate that these two agents reduce the amount of advanced glycosylation end products which form. The agents do not alter the formation of early glycosylation products.

EXAMPLE II

To more precisely measure the effect of the agents on the inhibition of protein crosslinking, an assay system was devised to measure the extent of *in vitro* binding of a soluble protein to an insoluble protein. This assay system mimics the events which occur in tissues in which serum proteins become bound to proteins in extravascular matrix and which lead to accumulation of protein and narrowing of vessel lumina in several tissues. These events *in vivo* give rise to kidney disease and atherosclerotic disease, and lead to the pathologies associated with diabetes and aging.

To measure protein trapping (i.e. binding or accumulation), gelatin (collagen) was coupled to activated agarose beads (Affigel 10, Bio-Rad Laboratories) by routine methods. After coupling, all remaining active sites on the beads were quenched by reaction with glycine ethyl ester.

The beads were incubated for two weeks with bovine serum albumin and 400 mM glucose-6-phosphate, a more reactive form of glucose which forms early glycosylation products with proteins more rapidly than does glucose. Also included in some experiments were the test agents, aminoguanidine, α -hydrazinohistidine, or lysine, at a concentration of 200 mM. The bovine serum albumin was radioiodinated so that the amount which became bound to the beads could be measured. The amount of radiolabel that became bound to the beads is a direct measure of protein trapping.

After a two-week incubation of the reaction mixtures at 37°C , the beads were washed extensively with chaotropic agents and the covalently bound radioactivity was determined. The results are set forth in Figure 2.

The left bar shows the control level of radiolabel incorporated into the beads in the absence of glucose-6-phosphate and in the absence of any test agents ("CONTROL COLLAGEN"). The second bar shows the high amount of incorporation in the presence of glucose-6-phosphate ("NEG. COLLAGEN"). This is likened to the presence of high concentrations of glucose in the blood of uncontrolled patients with diabetes and the pathological sequelae which result.

Figure 2 shows that the amount of protein trapping in the presence of either aminoguanidine ("NEG. COLLAGEN + I#2") or α -hydrazinohistidine ("NEG. COLLAGEN + I#1") is greatly reduced.

Lysine also reduced the amount of protein trapping to an extent similar to that of aminoguanidine (not shown). The results show the potential value of these compounds in vivo for reducing the trapping or soluble protein on to membranes and other tissues, and further evidence that these agents may be of value in reducing the pathogenesis of diabetes and aging.

EXAMPLE III

As a further evaluation of the compound aminoguanidine as a model for the prevention of protein trapping, crosslinking and formation of advanced glycosylation end products, the following experiment using calf skin collagen was performed. Collagen is a protein in the skin responsible for the suppleness of skin, and crosslinking leads to wrinkling, decreased elasticity, reduced susceptibility to proteolytic degradation, and other changes.

Collagen from samples of calf skin were extracted into acetic acid and then precipitated with 0.6 M sodium chloride. These procedures removed from the solution skin collagen that was already permanently crosslinked or denatured. Native collagen fibrils were reformed by dialysis against 0.02M phosphate buffer, and these were incubated for 3 weeks at 35°C in the presence of 140 mM glucose and with or without 200 mM aminoguanidine. After incubation, the samples were dialyzed and the degree of crosslinking was determined by two methods. First, the amount of reacted collagen which could be solubilized by treatment in 2% sodium dodecyl sulfate at 100°C was measured.

As shown in Figure 3A, collagen incubated with glucose and aminoguanidine was as soluble as collagen incubated in buffer alone. In contrast, collagen incubated in glucose without aminoguanidine was only 50% as soluble. This is further evidence that aminoguanidine has utility in the prevention of age-related changes in skin and other tissues.

The reacted collagen was further examined by cleaving it into fragments using cyanogen bromide treatment in formic acid. The resulting protein fragments were separated by size by sodium dodecyl sulfate - polyacrylamide gel electrophoresis. After electrophoreses, the protein fragments were identified in the gel using silver staining. The gel is shown in Figure 3B.

Referring to Figure 3B, Lane B contains collagen that was incubated with glucose alone. It is noted that a large amount of high molecular weight fragments form a continuous band at the top of the gel, indicating a large range of high molecular weight fragments. Some of this material could not enter the gel and is present in the 30% stacking gel above the gradient gel. Lane C contains the

collagen incubated with glucose and aminoguanidine, and it is noted that there is no large amount of high molecular weight material at the top of the lane, as all of the protein fragments separate well in the lower part of the gel. Lane A shows collagen incubated in PBS alone or held frozen during the incubation of the other samples, respectively. The far left lane is a series of molecular weight markers. Identical results were observed with and without the presence of disulfide bond reducing agents in the electrophoreses buffer.

The above data indicate that aminoguanidine reduces the amount of crosslinking which occurs when collagen is incubated with glucose, and suggest the utility of this agent for topical application to skin to prevent age-related changes, including loss of elasticity and wrinkling.

The above in vitro experiments all point to the value of aminoguanidine as an agent to inhibit the formation of advanced glycosylation end products which form in vitro from protein incubated in the presence of glucose or other reducing sugars. As glucose is present in the body and is elevated in amount in diabetes, and as proteins in the body are known to undergo crosslinking and form fluorescent compounds all indicative of advanced glycosylation end products, use of this agent in vivo would be useful in the prevention of the pathology associated with diabetes and changes that occur during aging.

Accordingly, the following experiment was performed to test the hypotheses of the present invention in an in vivo experiment.

EXAMPLE IV

To measure the level of advanced glycosylation end products in vivo, the kidneys of rats were examined for serum proteins which had become attached to glomerular basement membranes. This was determined to be a good model in which to study this process as it is known that significant kidney pathology occurs in untreated diabetes as a result of the buildup of extravasated plasma protein in the extravascular matrix in this organ.

The experiment consisted of giving both normal and diabetic rats daily intraperitoneal doses of the agent aminoguanidine hydrochloride at a dose of 25 mg per kilogram of body weight, for a period of 16 weeks. The hydrochloride salt of aminoguanidine was used as it is more soluble and less irritating than the parent free base compound. Diabetes was induced prior to drug therapy with a single dose of streptozotocin. Control animals, both diabetic and normal, received no drug.

At the end of the agent therapy, animals were sacrificed and the kidneys were removed. Each organ was removed from its capsule and the medulla was removed. The remainder of the tissue,

principally containing glomeruli, was frozen on dry ice and stored at -70°C . Tissue from five animals in each treatment group was combined for processing.

To prepare glomerular basement membranes, tissue was cut into slices and passed through a series of sieves (170, 100 and 270) to separate glomeruli from tubules and other undesired tissue elements as described (Beisswenger, P.J., Spiro R.G., *Diabetes*, 22:180-193, 1973). Glomerular purity was found to be 80-90%. The final material was collected and centrifuged at 1500 rpm for fifteen minutes to pellet the glomeruli, which were frozen at -70°C .

Thawed isolated glomeruli were then disrupted by treatment in a Branson Sonifier 200 cell disrupter for four one-minute intervals on ice with a one-minute rest between sonications. Samples were examined in a phase contrast microscope to ensure that all of the glomeruli were disrupted. Glomerular basement membranes were then pelleted by centrifugation at 3000 rpm for ten minutes, washed with 1 M sodium chloride followed by distilled water. The remaining pellet of purified glomerular basement membranes was frozen and lyophilized.

An enzyme immunoassay was used to measure the amount of serum immunoglobulin G (IgG) that became bound to the glomerular basement membranes of the normal and diabetic animals after treatment with and without the agent. To measure IgG, 6 mg samples of lyophilized glomerular basement membrane tissue was suspended in 0.5 mM of 0.05 M carbonate buffer, pH 7.6, and 0.5 mM of a 1:5,000 dilution of rat anti-IgG antibody conjugated to alkaline phosphatase (Dynatech Corp.) was added. The mixture was incubated overnight in polystyrene tubes which were preblocked by incubation for two hours in 3% goat serum plus 0.05% Tween \otimes 20 in phosphatase buffered saline (PBS), followed by two rinses in PBS plus Tween \otimes .

After overnight incubation to allow the antibody to bind to any IgG crosslinked to the glomerular basement membranes, the membranes were pelleted by centrifugation at 3200 rpm for five minutes and were washed free of the unbound antibody-enzyme conjugate with four rinses with PBS plus Tween \otimes followed by three rinses with distilled water. To measure the amount of antibody-enzyme conjugate remaining bound, 0.5 mM of substrate solution (containing 1 mg/mL paranitrophenyl-phosphate in 10% diethanolamine, pH 9.8), was added and incubations were carried out for 30 minutes at room temperature. The reaction was stopped with the addition of 0.2 mL of M sodium hydroxide, and the absorbance at 400 nm was measured.

Figure 4 shows the results of this experiment. As can be seen, diabetic animals had a high level of IgG bound to their glomerular basement membranes ("D") and normal animals had about one-fifth the amount ("N"). Diabetic animals which received daily doses of aminoguanidine hydrochloride showed the same low level of IgG in normal animals ("D + I"). Normal animals receiving the drug had about the same low level ("N + I").

These experiments indicated that aminoguanidine prevented the trapping and accumulation of this plasma protein on the rat glomerular basement membranes. Presumably, the trapping of this and other serum proteins in the kidney, eye, on artery walls, and in other tissues known to be affected from this pathology would likewise be reduced. Trapping of lipoproteins on artery walls is known to contribute to atherosclerotic disease.

These *in vivo* experiments provide further evidence from the *in vitro* experiments that this type of drug therapy has benefit in reducing the pathology associated with the advanced glycosylation of proteins and the formation of crosslinks between proteins and other macromolecules. Drug therapy may be used to prevent the increased trapping and crosslinking of proteins that occurs in diabetes and aging which leads to sequela such as arterial disease, including renal disease, hypertension, retinal damage, and extra-vascularly, damage to tendons, ligaments and other joints. This therapy might retard atherosclerosis and connective tissue changes that occur with diabetes and aging. Both topical, oral and parenteral routes of administration to provide therapy locally and systematically are contemplated.

Claims

1. A method for inhibiting the advanced glycosylation of an isolated quantity of target protein comprising contacting the target protein with an effective amount of a composition comprising an agent selected from the group consisting of aminoguanidine, α -hydrazinohistidine, amino acids, esters and amides of amino acids, and mixtures thereof, capable of reacting with a carbonyl moiety of an early glycosylation product resulting from the initial glycosylation of the target protein.
2. A method for inhibiting the advanced glycosylation of a target protein found in food-stuffs comprising contacting the target protein with an effective amount of the composition comprising an agent selected from the group consisting of aminoguanidine, α -hydrazinohistidine, amino acids, esters and

amides of amino acids, and mixtures thereof, capable of reacting with a carbonyl moiety of an early glycosylation product resulting from the initial glycosylation of the target protein.

3. Use of an agent selected from the group consisting of aminoguanidine, α -hydrazinohistidine, amino acids, esters and amides of amino acids, and mixtures thereof, capable of reacting with a carbonyl moiety of an early glycosylation product resulting from the initial glycosylation of the target protein for the manufacture of a pharmaceutical composition for treating diseases associated with advanced glycosylation of proteins, but not including the combination of α -hydrazinohistidine and diabetic disorders. 10
4. Use of aminoguanidine for the manufacture of a pharmaceutical composition for treating diseases associated with advanced glycosylation of proteins. 20
5. Use of lysine for the manufacture of a pharmaceutical composition for treating diseases associated with advanced glycosylation of proteins. 25
6. Use of α -hydrazinohistidine for the manufacture of a composition for treating diseases associated with advanced glycosylation of proteins, but not including diabetic disorders. 30
7. Use according to claims 3, 4, 5 or 6 wherein the target protein is selected from the group consisting of collagen, elastin, lens protein, blood vessel walls, nerve protein and glomerular basement membrane. 35
8. Use according to claims 3, 4, 5 or 6 wherein the pharmaceutical composition comprises a pharmaceutically acceptable carrier. 40
9. Use according to claims 3, 4, 5 or 6 wherein the pharmaceutical composition is adapted for administration parenterally. 45
10. Use according to claims 3, 4, 5 or 6 wherein the pharmaceutical composition is adapted for administration topically. 50
11. Use according to claims 3, 4, 5 or 6 wherein the pharmaceutical composition is adapted for administration orally. 55
12. Use according to claims 3, 4, 5, 6 or 11 wherein a pharmaceutical composition is prepared in a dosage form wherein the agent for treating is present in an amount calculated to

deliver a dose of up to 25 mg/kg body weight to a patient under treatment.

13. Use according to claims 3, 4, 5 or 6 wherein a pharmaceutical composition is prepared in an ointment form and the agent for treating is present in an amount of up to 10% by weight.

Patentansprüche

1. Verfahren zur Hemmung der fortgeschrittenen Glykosylierung einer isolierten Menge eines Zielproteins mit dem Schritt, daß das Zielprotein mit einer wirksamen Menge einer Zusammensetzung in Kontakt gebracht wird, welche einen Wirkstoff enthält aus der Gruppe Aminoguanidin, α -Hydrazinohistidin, Aminosäuren, Ester und Amide von Aminosäuren, und Mischungen davon, die befähigt sind, mit einem Carbonyl-Rest eines frühen Glykosylierungsprodukts aus der Anfangsphase der Glykosylierung des Zielproteins zu reagieren.
2. Verfahren zur Hemmung der fortgeschrittenen Glykosylierung eines in Nahrungsmitteln vorkommenden Zielproteins mit dem Schritt, daß das Zielprotein mit einer wirksamen Menge der Zusammensetzung in Kontakt gebracht wird, welche einen Wirkstoff aus der Gruppe Aminoguanidin, α -Hydrazinohistidin, Aminosäuren, Ester und Amide der Aminosäuren, und Mischungen davon enthält, welche befähigt sind, mit einem Carbonyl-Rest eines frühen Glykosylierungsprodukts aus der Anfangsphase der Glykosylierung des Zielproteins zu reagieren.
3. Verwendung eines Wirkstoffs aus der Gruppe Aminoguanidin, α -Hydrazinohistidin, Aminosäuren, Ester und Amide von Aminosäuren, und Mischungen davon, welche befähigt sind, mit einem Carbonyl-Rest eines frühen Glykosylierungsprodukts zu reagieren, welches aus der Anfangsphase der Glykosylierung des Zielproteins stammt, zur Herstellung einer pharmazeutischen Zusammensetzung zur Behandlung von mit fortgeschrittener Glykosylierung von Proteinen assoziierten Krankheiten, jedoch unter Ausschluß der Kombination von α -Hydrazinohistidin und diabetischer Störung.
4. Verwendung von Aminoguanidin zur Herstellung einer pharmazeutischen Zusammensetzung zur Behandlung von mit fortgeschrittener Glykosylierung von Proteinen assoziierten Krankheiten.
5. Verwendung von Lysin für die Herstellung einer pharmazeutischen Zusammensetzung zur

Behandlung von mit fortgeschrittener Glykosylierung von Proteinen assoziierten Krankheiten.

6. Verwendung von α -Hydrazinohistidin zur Herstellung einer Zusammensetzung für die Behandlung von mit fortgeschrittener Glykosylierung von Proteinen assoziierten Krankheiten, jedoch unter Ausschluß diabetischer Störungen.
7. Verwendung gemäß den Ansprüchen 3, 4, 5 oder 6 wobei das Zielprotein aus der Gruppe Kollagen, Elastin, Linsenprotein, Blutgefäßwänden, Nervenprotein und Glomerulus-Basalmembran ausgewählt ist.
8. Verwendung gemäß den Ansprüchen 3, 4, 5 oder 6, wobei die pharmazeutische Zusammensetzung einen pharmazeutisch verträglichen Träger umfaßt.
9. Verwendung gemäß den Ansprüchen 3, 4, 5 oder 6, wobei die pharmazeutische Zusammensetzung an parenterale Verabreichung angepaßt ist.
10. Verwendung gemäß den Ansprüchen 3, 4, 5 oder 6, wobei die pharmazeutische Zusammensetzung an topische Verabreichung angepaßt ist.
11. Verwendung gemäß den Ansprüchen 3, 4, 5 oder 6, wobei die pharmazeutische Zusammensetzung an orale Verabreichung angepaßt ist.
12. Verwendung gemäß den Ansprüchen 3, 4, 5, 6 oder 11, wobei eine pharmazeutische Zusammensetzung in einer Dosierungsform hergestellt ist, in welcher der Wirkstoff zur Behandlung in einer Menge vorliegt, welche so berechnet ist, daß eine Dosis bis zu 25 mg/kg Körpergewicht an einen unter Behandlung stehenden Patienten abgegeben wird.
13. Verwendung gemäß den Ansprüchen 3, 4, 5 oder 6, wobei eine pharmazeutische Zusammensetzung in Form einer Salbe hergestellt ist und der Wirkstoff zur Behandlung in einer Menge von bis zu 10 Gew.-% vorliegt.

Revendications

1. Méthode d'inhibition de glycosylation avancée d'une quantité isolée de protéine cible comprenant la mise en contact de la protéine cible avec une quantité efficace d'une composition comprenant un agent choisi dans le groupe

composé d'aminoguanidine, α -hydrazinohistidine, acides aminés, esters et amides d'acides aminés, et leurs mélanges, capables de réagir avec une demi-portion de carbonyle d'un produit antérieur de glycosylation résultant de la glycosylation initiale de la protéine cible.

2. Méthode d'inhibition de la glycosylation avancée d'une protéine cible trouvée dans des produits alimentaires comprenant la mise en contact de la protéine cible avec une quantité efficace de la composition comprenant un agent choisi dans le groupe consistant en aminoguanidine, α -hydrazinohistidine, acides aminés, esters et amides d'acides aminés, et leurs mélanges, capables de réagir avec une demi-portion de carbonyle d'un produit de glycosylation antérieure résultant de la glycosylation initiale de la protéine cible.

3. Utilisation d'un agent choisi dans le groupe composé de aminoguanidine, α -hydrazinohistidine, acides aminés, esters et amides d'acides aminés, et leurs mélanges, capables de réagir avec une demi-portion de carbonyle d'un produit de glycosylation antérieure résultant de la glycosylation initiale de la protéine cible pour la fabrication d'une composition pharmaceutique de traitement des maladies associées à la glycosylation avancée des protéines, mais n'incluant pas la combinaison de α -hydrazinohistidine et de troubles diabétiques.

4. Utilisation d'aminoguanidine pour la fabrication d'une composition pharmaceutique de traitement des maladies associées avec la glycosylation avancées des protéines.

5. Utilisation de lysine pour la fabrication d'une composition pharmaceutique de traitement des maladies associées à une glycosylation avancée des protéines.

6. Utilisation d' α -hydrazinohistidine pour la fabrication d'une composition pour traiter les maladies associées à la glycosylation avancée des protéines, mais n'incluant pas de troubles diabétiques.

7. Utilisation suivant les revendications 3, 4, 5 ou 6, dans laquelle la protéine cible est choisie dans le groupe composé de collagène, élastine, protéine de cristallin, parois de vaisseau sanguin, protéine nerveuse et membrane de soubassement glomérulaire.

8. Utilisation suivant les revendications 3, 4, 5 ou 6, dans laquelle la composition pharmaceuti-

que comprend un support acceptable pharmaceutique.

9. Utilisation suivant les revendications 3, 4, 5 ou 6, dans laquelle la composition pharmaceutique est adaptée à une administration parentérale. 5
10. Utilisation suivant les revendications 3, 4, 5 ou 6, dans laquelle la composition pharmaceutique est adaptée à une administration topique. 10
11. Utilisation suivant les revendications 3, 4, 5 ou 6, dans laquelle la composition pharmaceutique est adaptée à une administration orale. 15
12. Utilisation suivant les revendications 3, 4, 5, 6 ou 11, dans laquelle une composition pharmaceutique est préparée en une forme de dosage dans laquelle l'agent de traitement est présent en une quantité calculée pour délivrer une dose, qui va jusqu'à 25 mg/kg de poids de corps, destinée au patient en traitement. 20
13. Utilisation suivant les revendications 3, 4, 5 ou 6, dans laquelle une composition pharmaceutique est préparée en une forme d'onguent et l'agent de traitement est présent en une quantité allant jusqu'à 10 % en poids. 25

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Inhibition of Advanced Glycosylation Product Formation in vitro

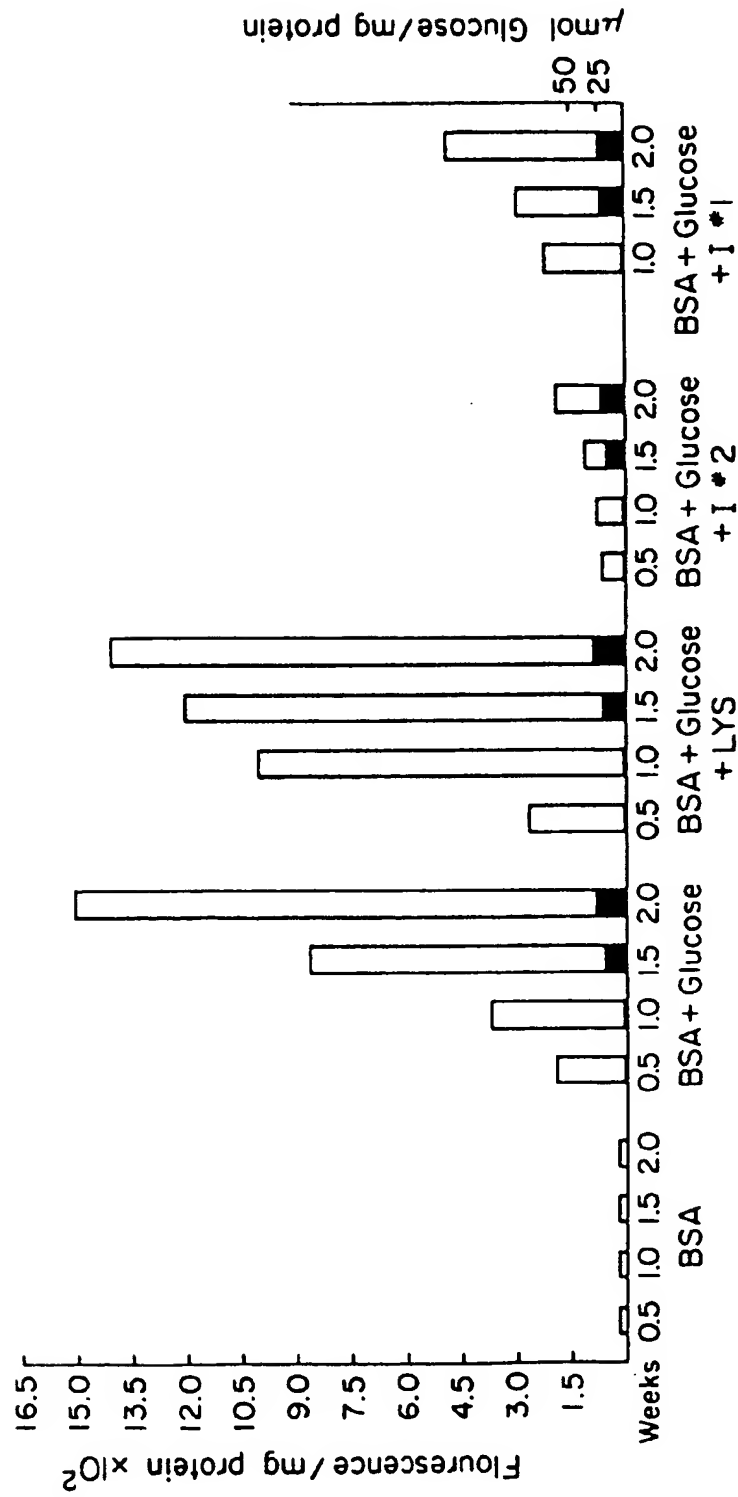


Fig. 1

FIG.2

Covalent Trapping of BSA by
Nonenzymatically Glycosylated Collagen

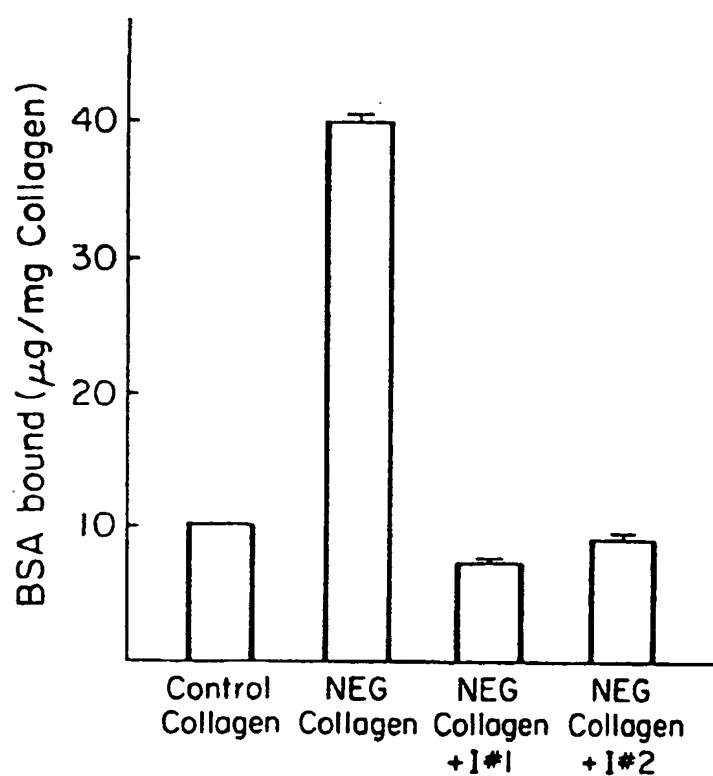
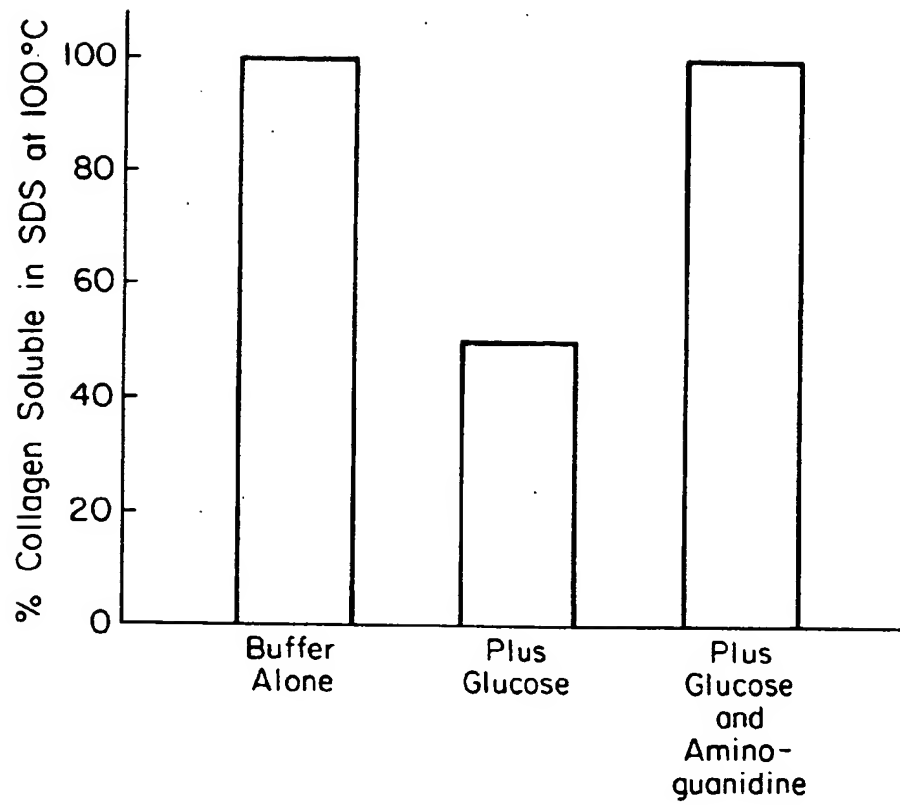


FIG.3A



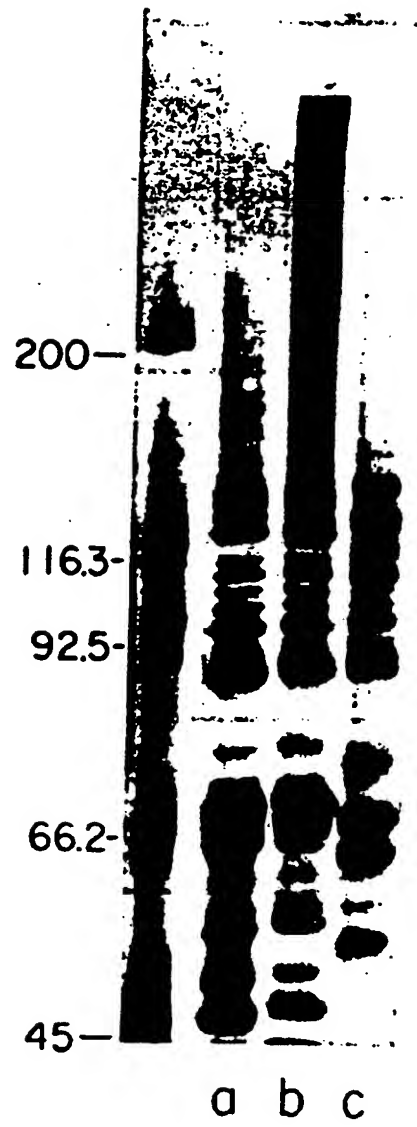
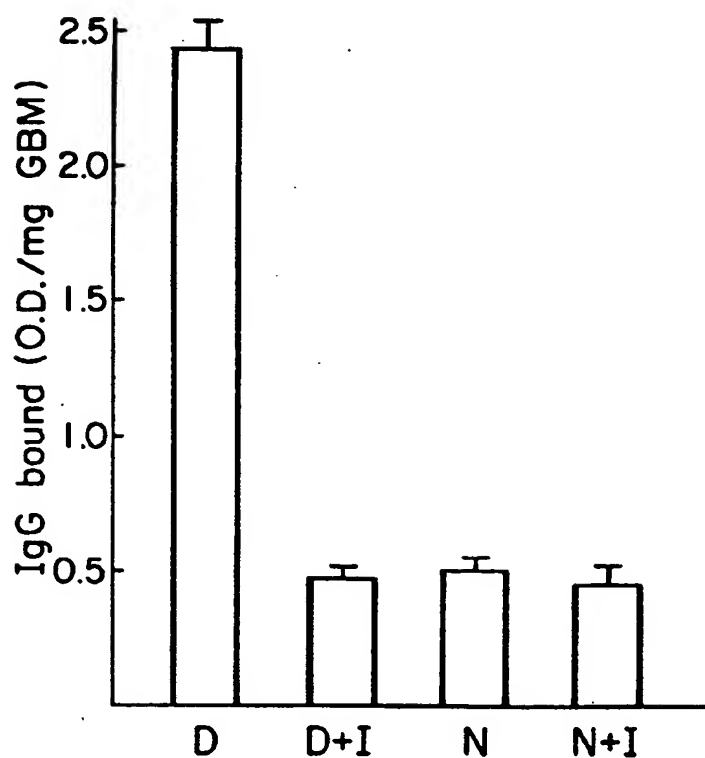


FIG.3B

FIG.4

Inhibition of Advanced Glycosylation
Product Formation in vivo



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